



Chapter 2. Global warming and breeding, biotechnology in aquaculture

Prof., Dr. Halyna Krusir

Prof., Dr. Maryna Mardar

Assoc. prof. Olha Sahdieieva

Odesa National University of Technology

Introduction

Aquatic ecosystems, essential to global biodiversity and human livelihoods, are undergoing unprecedented changes due to global warming. Rising temperatures, driven by anthropogenic climate change, are disrupting breeding cycles, survival rates, and the genetic integrity of aquatic species. Simultaneously, the field of aquaculture is navigating these challenges through innovative biotechnological solutions. With the world's reliance on aquatic resources increasing to meet food security demands, the integration of advanced breeding technologies, including selective breeding, genomic selection, and CRISPR/Cas9 gene editing, offers transformative potential to address the dual crises of climate change and sustainable aquaculture.

This chapter delves into the intricate interplay between environmental changes and biotechnological advances in aquaculture. It begins by exploring how global warming alters the breeding cycles and survival dynamics of aquatic species, causing significant shifts in population structures and ecosystem functions. The focus then shifts to groundbreaking biotechnological solutions, such as selective breeding and genomic selection, which are enhancing the resilience and productivity of aquaculture species. Furthermore, the revolutionary CRISPR/Cas9 gene-editing technology is discussed, highlighting its applications in improving disease resistance, growth rates, and environmental adaptability in various fish species. Finally, the ethical, environmental, and regulatory considerations surrounding these technologies are examined, emphasizing the need for sustainable and responsible innovation in aquaculture.

The implications of these discussions are far-reaching, influencing not only the future of aquaculture but also global efforts to mitigate biodiversity loss and climate change impacts. This introduction sets the stage for a comprehensive analysis of the challenges and opportunities presented by the integration of biotechnology into aquaculture under the shadow of global warming.

1. Impact of Global Warming on Aquatic Species Breeding

1.1. Changes in Breeding Cycles: Increased water temperatures can alter the breeding cycles of aquatic species, affecting spawning times, growth rates, and survival rates of larvae.

Global warming, driven by human-induced climate change, is having a profound impact on ecosystems around the world, including aquatic environments. One of the most significant areas affected by rising temperatures is the breeding of aquatic species. Changes in water temperature are altering breeding cycles, spawning times, growth rates, and the survival rates of offspring, leading



to shifts in the structure and functioning of aquatic populations. This literature review aims to explore how these environmental changes are influencing aquatic species, focusing on changes in breeding cycles and genetic adaptation.

Changes in Breeding Cycles. Rising water temperatures due to global warming are one of the primary drivers of changes in the reproductive behavior of aquatic species. Many species rely on specific temperature cues to initiate breeding. With increasing temperatures, the timing of breeding events has shifted, and these shifts can lead to mismatches between species and their habitats.

Spawning Times. Studies have shown that many aquatic species are breeding earlier in the year due to warmer water temperatures. For instance, fish species such as the Atlantic cod (*Gadus morhua*) and the European perch (*Perca fluviatilis*) have been observed to spawn earlier in the season in response to increased water temperatures (Tompkins et al., 2017). While earlier spawning may initially seem beneficial, it often leads to a mismatch with the availability of food resources for larvae, as phytoplankton, a primary food source for many young fish, may not be available at the same time (Durant et al., 2007). This could result in decreased survival rates of offspring, further impacting population dynamics.

Moreover, earlier spawning does not necessarily guarantee success, as species may spawn before conditions are optimal for larvae survival. The mismatch in timing can lead to a reduced number of viable offspring, potentially leading to long-term population declines (O'Reilly et al., 2008).

Growth Rates and Metabolic Effects. The increase in water temperature also influences the metabolic rates of aquatic species. Warmer temperatures typically accelerate the growth of many species by speeding up metabolic processes (Angilletta et al., 2004). However, this increase in growth rate may not always be beneficial. Species that grow too quickly in warmer waters may not develop the necessary size or strength to survive into adulthood, which can lead to weaker individuals with lower chances of successful reproduction (Heath et al., 2014). Additionally, faster growth does not always correlate with an increase in reproductive success, as the species may face a mismatch in the timing of their developmental milestones and environmental conditions.

Survival Rates of Larvae. The early life stages of aquatic species are often the most vulnerable to environmental changes, and rising water temperatures can further exacerbate these vulnerabilities. Elevated temperatures can reduce the oxygen levels in the water, affecting the survival rates of larvae, which require high oxygen concentrations for proper development (Pörtner et al., 2014). Furthermore, higher temperatures may stress juvenile organisms, leaving them less capable of handling other environmental challenges, such as predation or food scarcity (Walther et al., 2002).

1.2. Genetic Adaptation: Some species may adapt genetically to changing temperatures, while others may face reduced reproductive success or population decline.

While environmental changes are causing challenges for aquatic species, some have the potential to adapt genetically to the changing conditions. Genetic adaptation involves changes in the genetic makeup of populations over time that allow species to cope with environmental stressors, including higher temperatures.

Adaptation to Temperature Changes. Research has suggested that certain species have shown some degree of genetic adaptation to rising temperatures. For example, studies on the Atlantic cod have found evidence of local adaptation to varying thermal conditions in different geographic areas (Jorgensen et al., 2017). Some populations of cod that live in warmer waters have developed genetic



traits that allow them to spawn successfully at higher temperatures. Similarly, some fish species may exhibit shifts in their reproductive timing or physiological tolerance, adapting to warmer environments over multiple generations (Lynch et al., 2014).

However, the ability of species to adapt genetically is limited by factors such as genetic diversity and the speed at which environmental changes occur. Species with low genetic diversity or those in rapidly warming habitats may struggle to adapt quickly enough to avoid population declines (Fischer et al., 2014). Additionally, the process of genetic adaptation is slow, and the rate of warming may exceed the ability of some species to genetically adapt in a timely manner.

Reduced Reproductive Success and Population Decline. While some species may successfully adapt to warming temperatures, others may face challenges that reduce their reproductive success or lead to population declines. For instance, species with specialized breeding requirements, such as those that rely on very specific temperature ranges for spawning, may find it difficult to cope with the rapid temperature shifts caused by global warming (Parmesan, 2006). In such cases, reproductive success may decrease, and populations may experience a decline in numbers or even local extinctions.

Species that do not adapt genetically to rising temperatures may be unable to reproduce successfully in their native habitats, leading to a loss of genetic diversity and further reducing their chances of survival in the face of climate change (Chevin et al., 2010).

The impact of global warming on aquatic species breeding is multifaceted, involving shifts in breeding cycles, changes in growth rates, and alterations in the survival rates of offspring. Rising temperatures have led to earlier spawning in many species, but this may cause a mismatch with food availability and optimal environmental conditions, resulting in lower survival rates for larvae. While some species may be able to genetically adapt to changing temperatures, the rate of environmental change may exceed their ability to do so, leading to reduced reproductive success and potential population declines. Further research is necessary to understand the long-term consequences of these changes on aquatic ecosystems and to develop strategies for mitigating the effects of climate change on these species.

2. Biotechnological Advances in Aquaculture Breeding

2.1 Selective Breeding: Use of selective breeding techniques to develop strains of fish and shellfish that are more resilient to higher temperatures and other climate-related stresses.

Aquaculture is a rapidly growing sector, contributing significantly to global food security. As the climate continues to change, aquaculture faces increasing challenges, such as rising temperatures and more frequent extreme weather events. To address these challenges, biotechnological advancements, particularly in selective breeding and genomic selection, are being increasingly applied to develop aquaculture species that are more resilient to climate-related stresses.

Selective Breeding

Selective breeding has been a cornerstone of aquaculture for decades, helping to enhance the productivity and resilience of farmed species. The process involves selecting individuals with desirable traits for reproduction, thus gradually improving the genetic composition of populations. Traditional selective breeding in aquaculture has focused on traits such as growth rate, disease resistance, and feed conversion efficiency. With climate change intensifying environmental



stressors, there is a growing emphasis on breeding for traits that confer greater resilience to elevated water temperatures and other climate-related challenges.

Research has shown that selective breeding can help aquaculture species, such as fish and shellfish, adapt to warmer environments. For example, studies on the Atlantic salmon have shown that selective breeding can enhance heat tolerance, potentially enabling farmed populations to survive in warmer waters that result from climate change (Gjøen et al., 2018). Additionally, selective breeding programs are increasingly focusing on traits like disease resistance and the ability to withstand hypoxic conditions, which are likely to become more prevalent as water temperatures rise (Houston et al., 2018).

Selective breeding for climate resilience also includes enhancing behavioral traits. For instance, fish that exhibit greater tolerance to stressors such as crowding and handling can better withstand the harsher conditions created by climate change (Huntingford et al., 2020). These breeding programs aim to ensure that aquaculture species can continue to thrive under a changing climate, contributing to long-term sustainability.

2.2 Genomic Selection: Implementation of genomic tools to identify and propagate desirable traits, enhancing the ability of aquaculture species to thrive in a changing climate.

The use of modern biotechnology to enhance production of aquatic species holds great potential not only to meet demand but also to improve aquaculture. Genetic modification and biotechnology also holds tremendous potential to improve the quality and quantity of fish reared in aquaculture. There is a growing demand for aquaculture; biotechnology can help to meet this demand. As with all biotech-enhanced foods, aquaculture will be strictly regulated before approved for market. Biotech aquaculture also offers environmental benefits. When appropriately integrated with other technologies for the production of food, agricultural products and services, biotechnology can be of significant assistance in meeting the needs of an expanding and increasingly urbanized population in the next millennium. Successful development and application of biotechnology are possible only when a broad research and knowledge base in the biology, variation, breeding, agronomy, physiology, pathology, biochemistry and genetics of the manipulated organism exists. Benefits offered by the new technologies cannot be fulfilled without a continued commitment to basic research. Biotechnological programs must be fully integrated into a research background and cannot be taken out of context if they are to succeed.

Figure 2.1 shows the role of biotechnology in enhancing fish production.

Genomic selection, which uses genomic tools to identify and propagate desirable traits, represents a major leap forward in aquaculture breeding. This technique involves associating genetic markers with traits of interest, allowing for more efficient selection. Genomic selection can accelerate breeding programs by enabling breeders to identify individuals with the best genetic potential for resilience to environmental stressors.

One of the most promising applications of genomic selection in aquaculture is in improving heat tolerance. A study on the rainbow trout (*Oncorhynchus mykiss*) found that genomic selection could be used to identify markers linked to heat tolerance, enabling the development of strains that are better equipped to survive in warmer waters (Liu et al., 2020). By applying genomic selection to breeding programs, aquaculture species can be genetically tailored to thrive in environments that are expected to experience higher temperatures due to climate change.

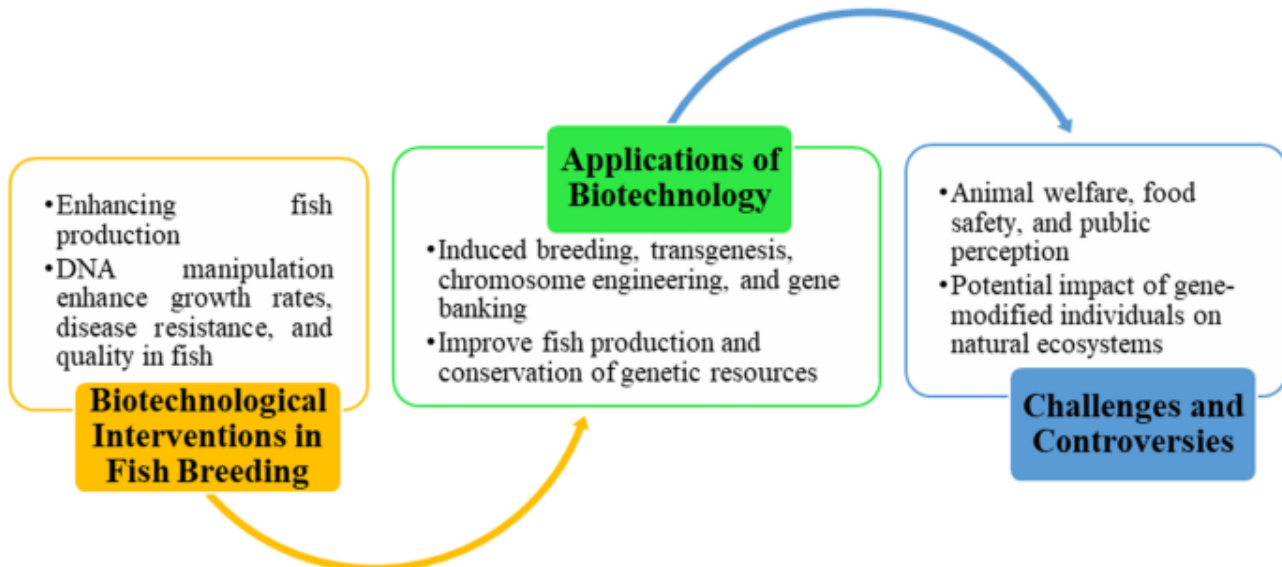


Figure 2.1. The role of biotechnology in enhancing fish production (Yang et al., 2021).

In addition to heat tolerance, genomic selection is being used to improve other climate-related traits, such as disease resistance and the ability to survive in low-oxygen environments. For example, genomic tools are being employed to identify genetic markers associated with resistance to the pathogen *Vibrio anguillarum*, which poses a significant threat to aquaculture species in warmer waters (Vázquez et al., 2018). By using genomic selection to breed fish that are more resistant to diseases, aquaculture systems can become more sustainable and less reliant on antibiotics, which are increasingly under scrutiny due to their environmental impact.

Genomic selection is also being integrated with traditional selective breeding to maximize genetic gain. The combination of genomic information with phenotypic data allows breeders to make more informed decisions about which individuals to select for reproduction. For example, genomic data can be used to predict the future performance of offspring, helping to avoid issues such as inbreeding and ensuring the long-term genetic health of aquaculture populations (Gjøen et al., 2018).

Integrating Selective Breeding and Genomic Selection

The integration of selective breeding and genomic selection is seen as a powerful strategy to ensure the resilience of aquaculture species in the face of climate change. Selective breeding provides a solid foundation by improving traits such as growth rate and disease resistance, while genomic selection accelerates the process and enhances the precision of breeding programs. Together, these techniques enable the rapid development of strains that are better suited to the changing environmental conditions.

In the case of Atlantic salmon, for example, both selective breeding and genomic selection have been employed to create strains that are more resistant to higher temperatures and diseases (Gjøen et al., 2018). The combination of these two approaches has the potential to significantly increase the sustainability of aquaculture by developing strains that can thrive in warmer, more variable environmental conditions.

Challenges and Future Directions

While biotechnological advances hold great promise for improving aquaculture breeding, there are challenges that need to be addressed. One of the key concerns is the potential for genetic



homogenization in farmed populations, which can lead to inbreeding depression and reduced genetic diversity. It is crucial for breeding programs to manage genetic diversity effectively, ensuring that aquaculture species remain adaptable to future environmental changes (Houston et al., 2018).

Moreover, the implementation of genomic selection requires significant investment in genomic resources, including the development of high-quality reference genomes and genetic markers. While genomic tools have become more accessible in recent years, the cost and complexity of these tools remain a barrier for some aquaculture industries (Huntingford et al., 2020).

Despite these challenges, the continued development of genomic technologies, combined with advances in computational tools and breeding strategies, holds great potential for improving the resilience of aquaculture species to climate change.

Conclusion

Biotechnological advances in aquaculture breeding, including selective breeding and genomic selection, offer promising solutions to the challenges posed by climate change. By enhancing the resilience of aquaculture species to rising temperatures, disease, and other environmental stressors, these technologies can help ensure the sustainability of the industry. The integration of genomic selection with traditional breeding approaches is likely to be a key strategy for developing more climate-resilient strains of fish and shellfish. As the aquaculture sector continues to face the pressures of climate change, these biotechnological innovations will play a critical role in ensuring that aquaculture remains a viable and sustainable source of food for the global population.

3. Genetic Engineering and CRISPR

3.1. Genetic Engineering in Aquaculture

The use of biotechnological methods to improve the well-being of cultured organisms, increase productivity, and protect aquatic ecosystems has yielded encouraging results. Vaccines and immunostimulants, probiotics, prebiotics, symbiotics, paraprobiotics, phage treatment, antimicrobial peptides, gene therapy, RNA interference, and other biotechnological therapies are among them. Genetic advancements in aquaculture play a crucial role in increasing productivity, reducing production costs, and minimizing the environmental impact.

Examples of methods for editing fish genomes include CRISPR-Cas9, transcription activator-like effector nucleases, and zinc-finger nucleases. Molecular biology and transgenesis, gene banking, chromosome manipulation, hormonal treatments, raising fish with one or more parents, creating fish with different numbers of cells (polyploid, triploid, haploid, gynogenetic, and androgenetic), and the use of synthetic hormones in fish breeding are other methods used in fish biotechnology.

Innovations in biotechnological technologies have revolutionized fish genetic breeding, leading to significant advancements in the aquaculture industry (Yang et al., 2021).

Techniques such as genetic engineering and CRISPR-Cas9 have enabled the precise modification of fish genomes, resulting in strains with increased growth rates, disease resistance, and improved feed conversion efficiency. Selective breeding programs have been optimized through marker-assisted selection, allowing for the identification and propagation of desirable genetic traits more efficiently. Furthermore, reproductive technologies, including hormone-induced spawning and cryopreservation of gametes, have enhanced breeding success and genetic diversity. These biotechnological advancements have contributed to more sustainable and productive fish farming practices, meeting the increasing global demand for seafood. These tools play a crucial role in

avoiding the extinction of endangered fish species and improving commercial fish production. Additionally, other biotechnological methods, such as synthetic hormone use, monosex production, and transgenesis, contribute to fish breeding advancements. These tools play major roles in preventing the extinction of endangered fish species and enhancing commercial fish production. Additionally, other biotechnological methods, such as the use of synthetic hormones, monosex production, and transgenesis, significantly contribute to advancements in fish breeding. Figure 2.2 shows the various biotechnological innovations in fish breeding (Sankaran & Mandal, 2024).

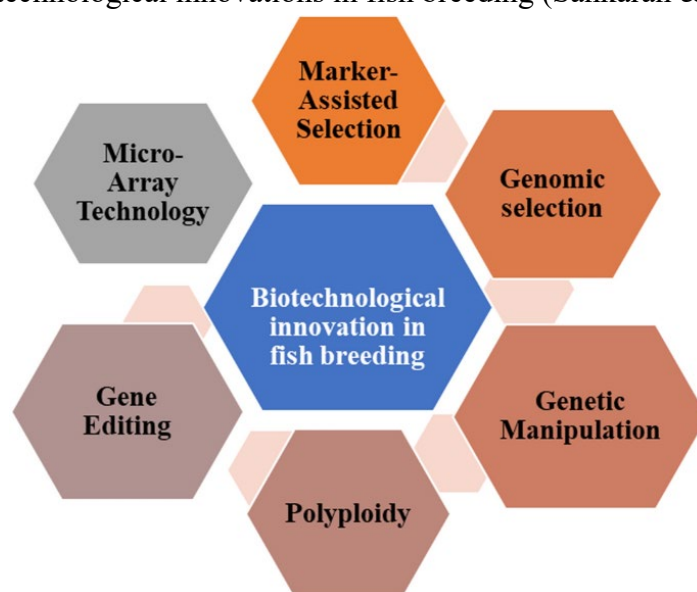


Figure 2.2. Biotechnological innovations in fish breeding

Genetic diversity represents a substantial resource that can be utilized to initiate selective breeding programs, which are proven to significantly improve the performance of the aquaculture sector. Facilitating the transfer of training and technology across different aquaculture sectors can greatly benefit lower-value species, enhancing their productivity and sustainability.

An organism’s genome can be modified by inserting synthetic DNA made from various sources using a process known as recombinant DNA technology. Implanting a genetic fragment containing our target gene into an existing genome is the first step in the procedure. In this technique, restriction enzymes, vectors, and host cells are utilized as tools. A variety of enzymes are involved in the processes of cutting, synthesizing, and binding. Enzymes such as restriction enzymes are part of this group. To transport and incorporate target genes, vectors are useful. The applications of recombinant DNA technology include gene cloning, gene therapy, and agriculture. Figure 2.3 shows the various steps involved in recombinant DNA technology (Sankaran & Mandal, 2024).

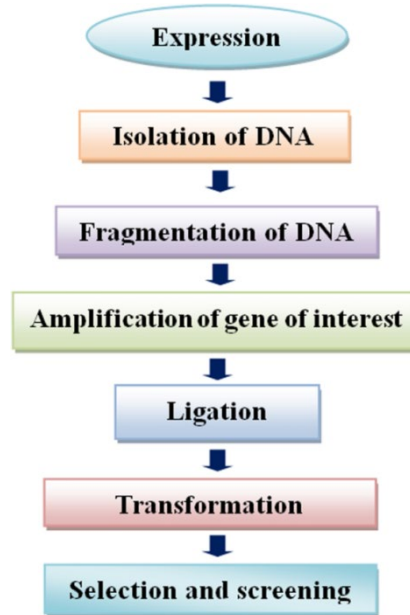


Figure 2.3 Main steps involved in recombinant DNA technology (Sankaran & Mandal, 2024)

3.2. CRISPR in Aquaculture

CRISPR/Cas9 represents a revolutionary tool in genetic engineering, enabling precise and targeted modifications of fish DNA to enhance traits such as pigmentation, growth, muscle quality, and disease resistance. This technology surpasses traditional breeding methods by offering a cheaper, easier, and more accurate approach to genetic enhancement. Its applications include improving growth performance (e.g., body weight, length, and muscle development), muscle quality, disease resistance, and sex determination. Furthermore, CRISPR/Cas9 provides promising solutions for increasing disease resistance by targeting immune-related genes and pathogen recognition pathways, reducing the need for antibiotics and chemical treatments. This technology has significantly advanced aquaculture by genetically optimizing key traits in fish species. For example, researchers have successfully manipulated germ cells in Atlantic salmon to control reproductive cell differentiation, improved feed conversion efficiency for growth in yellow catfish, achieved targeted gene modifications in tilapia, and minimized unintended off-target effects (Zhu et al., 2024).

Figure 2.4 shows the methods of CRISPR/Cas9 involved in gene editing. The Cas9 enzyme and guide RNA are the two main parts of the system. A streamlined variant of the CRISPR–Cas9 antiviral defense system found in bacteria serves as the basis for the CRISPR–Cas9 system. In vivo gene editing is made possible by inserting a synthetic guide RNA (gRNA) complexed with the Cas9 nuclease into a cell and then cutting the genome at a specific location. Because it allows for easy, affordable, and precise editing of genomes in vivo, this method is profoundly important in biotechnology and medicine. In addition to its potential utility in pest and disease management, it has other potential applications in the development of novel agricultural products, genetically modified organisms, and pharmaceuticals. Additionally, it shows promise in the management of hereditary disorders and disorders caused by somatic mutations, including cancer. The CRISPR/Cas9 system offers a simple RNA-guided method for causing targeted alterations at specific sites. Some phenotypes, including eye color or susceptibility to disease, can be induced by these DNA alterations. The system employs RNA molecules designed to match target DNA sequences in conjunction with the Cas9 nuclease enzyme.

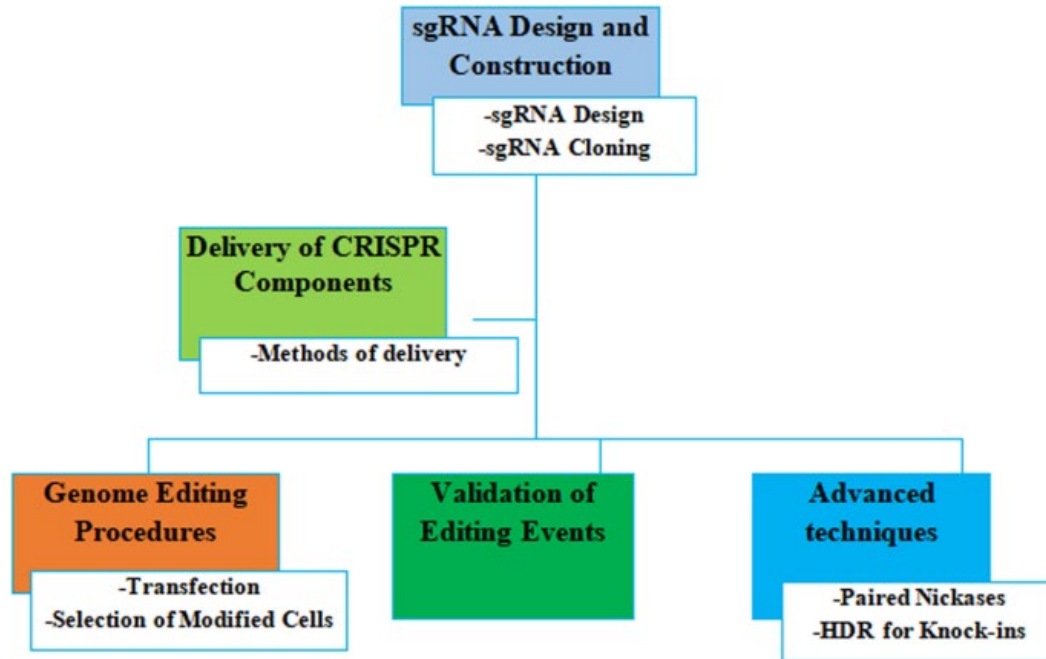


Figure 2.4. CRISPR/Cas9 gene editing (Sankaran & Mandal, 2024)

Although CRISPR/Cas9 has the potential to revolutionize the field of genetic engineering, it is not without its limitations. The accuracy of genome editing is a significant concern, as it results in permanent alterations to the genome. Additionally, its use in human germline genetic modification is highly controversial. In general, the utilization of CRISPR/Cas9 gene editing has the capacity to revolutionize the fields of biotechnology and medicine. However, exercising prudence and contemplating the ethical ramifications associated with its application are crucial (Sankaran & Mandal, 2024).

3.2.1. Disease resistance

Disease resistance is a critical trait in aquaculture, reflecting a species' ability to withstand infections, poor water quality, and environmental changes. CRISPR/Cas9-mediated genome editing has emerged as a powerful method to enhance this resistance. By integrating antimicrobial peptide genes (AMGs) into fish genomes, CRISPR/Cas9 reduces bacterial colonization, increases survival post-infection, and alters immune-related gene expression. This precision editing has led to significant advances, such as enhancing resistance to infectious pancreatic necrosis (IPN) and bacterial cold-water disease in salmon and targeting the JAM-A gene in grass carp to block viral entry, conferring immunity to grass carp reovirus (GCRV).

In tilapia, CRISPR/Cas9 has edited genes linked to immune responses, improving resistance to bacterial pathogens like *Streptococcus agalactiae* and *Aeromonas hydrophila*. Similarly, in catfish, this technology has targeted immune-regulating genes, increasing survival rates post-pathogen exposure. These advances have been complemented by knock-in techniques to introduce foreign genes, improving disease resistance while enhancing growth and nutritional value in species like tilapia and catfish (Zhu et al., 2024).

3.2.2. Fish growth and muscle quality

CRISPR/Cas9 has been instrumental in improving growth rates and muscle quality across aquaculture species, including Nile tilapia, channel catfish, common carp, and rainbow trout. By targeting growth hormone-related genes such as *myostatin* (*mstn*), which inhibits muscle growth,



researchers have achieved significant enhancements in body mass and muscle development. For instance, channel catfish with disrupted *mstn* genes showed a 29.7% increase in body weight, while similar modifications in olive flounder and red sea bream boosted muscle mass and optimized commercial fish size.

Beyond growth, CRISPR/Cas9 enables the study of developmental processes and human disease models using zebrafish, a widely used organism for genetic research. Transgenic techniques have further advanced fish farming by overexpressing growth hormone genes in species like Atlantic salmon, achieving rapid growth and higher yields to meet global protein demand. These genetic modifications, combined with optimized nutrition and selective breeding, enhance muscle texture and overall aquaculture efficiency (Zhu et al., 2024).

3.2.3. Off-Target Effects in CRISPR/Cas9

While CRISPR/Cas9 offers unparalleled precision, off-target effects remain a concern. These unintended edits can affect non-target genome regions, potentially causing adverse effects. Recent advancements, including high-fidelity Cas9 variants (e.g., SpCas9-HF1, eSpCas9), have significantly reduced off-target activity. Improved guide RNA (gRNA) design and algorithms like CRISPR-DO have enhanced specificity. Additionally, novel tools like base and prime editors allow precise genome modifications without inducing double-strand breaks, minimizing off-target mutations. Advanced delivery systems, such as nanoparticles and viral vectors, further increase accuracy in gene editing applications.

Zebrafish and other aquaculture species, including tilapia and Atlantic salmon, have benefited from these advancements. High-fidelity editing has enabled researchers to enhance growth, disease resistance, and other traits while maintaining genomic integrity.

The mechanism used by CRISPR/Cas9 in knockout genes in different fish species are indicated in Figure 2.5.

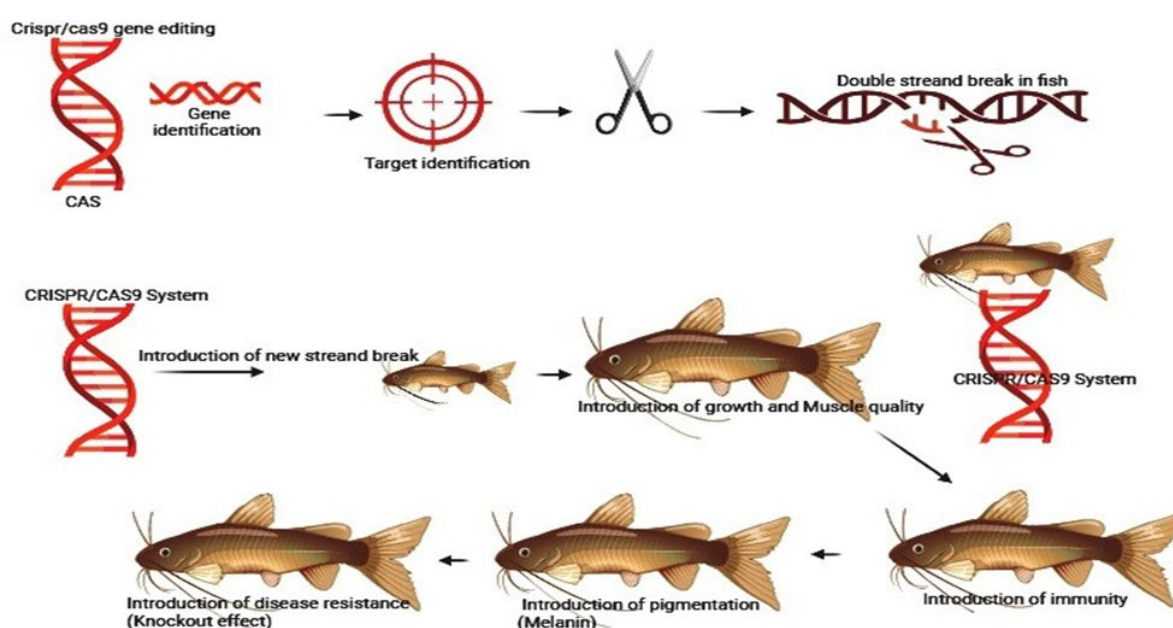


Figure 2.5. Steps of the application of CRISPR/Cas9 in aquaculture (First, a specific gRNA is designed to match the target gene sequence. Then, the Cas9 protein binds to the target DNA, causing a double-strand break. Finally, the break is repaired) (Zhu et al., 2024)



3.2.4. Sex Determination

Sex determination in fish involves genetic, environmental, and epigenetic factors, making it a complex but vital area of study in aquaculture. CRISPR/Cas9 has illuminated mechanisms of sex differentiation by precisely targeting relevant genes. For example, editing the *amh* gene in Nile tilapia resulted in phenotypic females from genetic males, demonstrating the gene's role in male sex determination. Similar studies in zebrafish have explored genes like *dmrt1* and *sox9a*, revealing the polygenic nature of sex determination in this species. Additionally, genome-wide CRISPR/Cas9 screenings have identified master regulators such as the *sdY* gene in rainbow trout, advancing our understanding of sex differentiation.

3.2.5. Effects of Using CRISPR/Cas9 in Gene Editing on Different Fish Species

CRISPR/Cas9 addresses challenges like disease outbreaks, poor growth rates, and environmental degradation in aquaculture. Its applications extend to controlling invasive species, engineering microorganisms for environmental remediation, and creating genetically modified fish for sustainable production. Genome editing offers solutions for enhancing fish traits while mitigating aquaculture's ecological footprint. For example, transgenic fish with improved feed conversion efficiency reduce resource use, supporting environmentally friendly practices.

By enabling precise genetic modifications, CRISPR/Cas9 has transformed aquaculture, paving the way for sustainable and efficient fish farming practices. Continued advancements in editing techniques, ethical considerations, and environmental management will further optimize its application in the industry. Table 2.1 provides a summary of the traits most commonly targeted for genome editing in fish aquaculture (Blix et al., 2021).

Table 2.1. Effects of CRISPR/Cas9 on Biological and Environmental Aspects of Fish Species

Applicable Fields	Impacts
Disease resistance	It is used to reduce the viral hemorrhagic septicemia virus (VHSV) infection of olive flounder hirame natural embryo (HINAE) cells.
	It enables gene editing in fish species such as salmon, tilapia, and shrimp to increase their resistance to diseases.
	It helps in the deletion of the <i>JAM-A</i> gene in grass carp cells, which significantly enhances resistance to grass carp reovirus (GCRV) infection.
	It helps enhance fish cell lines for host response and genetic resistance against infectious diseases, using Atlantic salmon and rainbow trout as model systems in aquaculture.
Environmental adaptation	It helps to edit genes in fish species, such as farmed salmon, to adapt to changing environments
Improved growth rates and muscles	It increases muscle growth by knocking out melanocortin (<i>mc4r</i>) receptor genes and has been experimentally tried on channel catfish and medaka fish.
	It improved the growth rates and increased muscle mass of the channel catfish by modifying the myostatin gene in channel catfish embryos.
	It helps increase the muscle mass of blunt snout bream due to the disruption of the <i>mstna</i>

CRISPR/Cas9 gene-editing technology has revolutionized aquaculture by enabling precise genetic modifications to improve traits such as disease resistance, growth, and sustainability. This tool also facilitates gene drives, increasing the inheritance rate of engineered genes to nearly 100%, accelerating the spread of desirable traits within populations.



Li et al., 2021 used CRISPR/Cas9 to create sterile, all-male Nile tilapia populations, resulting in faster growth rates and reduced ecological risks from escaped farmed fish. Similarly, Wargelius et al. enhanced disease resistance in Atlantic salmon by modifying genes essential for viral infection, addressing significant losses in aquaculture caused by pathogens like IPNV and SAV.

Other studies have leveraged CRISPR/Cas9 to enhance disease resistance in carp, tilapia, and catfish by targeting immune-related genes or pathogen recognition pathways. Growth-related gene editing has also yielded notable successes, such as *myostatin* knockouts in common carp, channel catfish, and red sea bream, leading to increased body size and growth rates.

CRISPR/Cas9's applications extend beyond production traits, enabling the creation of novel phenotypes. Examples include albino Nile tilapia and modified Pacific oysters with enhanced growth. The technology's versatility also spans species such as ridgetail shrimp, further demonstrating its transformative potential in aquaculture (Table 2.2).

Table 2.2. Applications of CRISPR/Cas9 in Various Fish Species and Their Impacts
(Zhu et al., 2024)

Fish Species	Technological Impacts
<i>Nile tilapia</i>	It is used to produce sterile Nile tilapia populations, reducing the risk of environmental damage from escaped fish.
<i>Atlantic salmon</i>	It helps in gene editing to create species that are highly resistant to viral infections, e.g. salmon
<i>Zebrafish</i>	It allows scientists to study mutations and genetic variants in zebrafish. It can be used to successfully integrate composite tags into zebrafish embryos, enabling precise labeling and visualization of cellular structures or proteins. This offers potential for studying protein dynamics, gene expression, and other biological processes in this model organism.
<i>Rainbow trout</i>	It has been shown to reduce the expression of the <i>igfbp-2b</i> gene in rainbow trout, influencing growth and development, but its impact on overall performance and the endocrine system remains unclear.
<i>Atlantic salmon and Rainbow trout</i>	It has been used to target unique genes associated with growth and immunity in Atlantic salmon, rainbow trout, and coho salmon cells.
<i>Japanese medaka</i>	It has the potential to increase muscle growth and body weight in farmed fish species such as medaka. However, further investigation is needed to determine its impact on production yield and fish health.
<i>Olive flounder</i>	It can be used to disrupt the <i>myostatin</i> gene in olive flounder, potentially increasing body weight and muscle tissue, but further research is needed to understand its effects on production efficiency and fish health
<i>Channel catfish</i>	It has been used to modify the <i>myostatin</i> gene in Channel catfish to improve muscle growth and quality, but further research is needed to fully understand its effects.

4. Cryopreservation and Assisted Reproductive Technologies

4.1. Aquaculture and Cryopreservation

Fish breeding is influenced by a variety of factors, and even the most experienced hatchery operators often encounter partial or complete failures in the breeding process. To achieve the desired quantity of seeds, induced breeding is widely regarded as an effective method. This approach facilitates the maturation and spawning of fish under unfavorable conditions, such as inadequate rainfall or extreme climate scenarios. However, repeated breeding efforts can take a significant toll on the health of broodstock within their limited lifespan. Replacing broodstock is



challenging due to logistical and physiological issues related to their transport. Consequently, the transportation of gametes has emerged as a promising alternative, offering benefits similar to those observed in animal husbandry.

The integration of biotechnological tools into fish breeding programs is essential for ensuring consistent and sustainable seed production. Cryopreservation presents a viable solution for producing high-quality seeds and genetically superior fish varieties. Recognizing its potential, the Food and Agriculture Organization (FAO) has identified cryopreservation as a critical strategy for conserving fish genetic resources (Betsy et al., 2022).

Cryopreservation refers to the preservation of biological samples at extremely low temperatures, effectively arresting metabolic activities and preserving structural and functional integrity for indefinite periods. This technology has become a cornerstone of reproductive biology, offering critical benefits for livestock and aquaculture industries. By maintaining temperatures below -130°C , metabolic activities cease entirely, enabling biological samples, such as cells, tissues, and even whole organisms, to remain viable upon thawing. Cryopreservation holds particular importance for preserving valuable genetic material, enhancing breeding programs, and supporting biodiversity conservation efforts (Fletcher & Rise, 2012).

4.2. Basic Principles of Cell Cryopreservation

4.2.1. Mechanisms of Preservation

Cryopreservation enables the preservation of gametes for extended periods, often spanning several years, without significantly affecting their fertilization capacity. By lowering the temperature to approximately -196°C , all biological and biochemical activities are halted, preventing processes that lead to cell death and DNA degradation. This technique is a powerful tool for supporting the long-term sustainability of aquaculture and biodiversity conservation.

However, ice formation within biological systems presents a significant challenge, as it can lead to mechanical damage and osmotic imbalance. Controlled cooling processes ensure that ice forms extracellularly, thereby creating a concentration gradient that facilitates water efflux from cells. This process prevents lethal intracellular ice formation. Advances in cryoprotective agents (CPAs) have been pivotal in mitigating these damages, allowing for the successful preservation of diverse cell types, tissues, and small biological structures. By refining the interplay between cooling rates, CPA concentrations, and cell-specific characteristics, researchers have enhanced cryopreservation outcomes.

4.2.2. Cryoprotective Agents

Cryoprotective agents play an essential role in reducing intracellular ice formation and maintaining protein and membrane integrity during freezing and thawing. These agents fall into two categories: permeable and non-permeable. Permeable CPAs, such as DMSO, glycerol, and methanol, penetrate the cell membrane to balance intracellular and extracellular osmotic pressures. Non-permeable CPAs, including sugars and certain proteins, primarily act extracellularly to modify the solution's freezing point and provide additional protection. Despite their benefits, CPAs must be used cautiously, as they can induce toxicity, osmotic stress, and chromosomal abnormalities if applied improperly. Balancing protective effects and potential adverse outcomes is a critical area of ongoing research.



4.2.3. Cooling and Thawing Protocols

The success of cryopreservation largely depends on the precise control of cooling and thawing protocols. Controlled freezing rates, typically ranging from $-40^{\circ}\text{C}/\text{min}$ to slower rates, are essential for minimizing ice crystal formation. Specialized biofreezers and nitrogen vapor methods are widely used to achieve these controlled conditions. Conversely, thawing must be rapid to prevent ice recrystallization, which can severely damage cellular structures. Emerging technologies, including programmable freezing devices and advanced thawing techniques, aim to standardize and optimize these processes for various biological materials, thereby improving survival rates and functional recovery (Fletcher & Rise, 2012).

4.3. Gamete Cryopreservation

4.3.1. Sperm Cryopreservation

Sperm cryopreservation represents one of the most successful applications of cryobiology, with well-established protocols in livestock and expanding applications in aquaculture. However, fish sperm exhibit significant differences from mammalian sperm, necessitating unique approaches. Key characteristics of fish sperm include their immotility in seminal plasma, activation of motility upon exposure to activating solutions, high sensitivity to osmotic changes, and relatively low ATP production. These unique traits underscore the need for tailored cryopreservation strategies to ensure viability and functionality upon thawing.

Developing effective protocols for fish sperm cryopreservation involves several critical steps:

Sperm Collection: Obtaining high-quality sperm free from contaminants is essential. Techniques such as abdominal massage, aspiration, or direct extraction from the testes are commonly employed, depending on the species. Care must be taken to avoid contamination with substances like urine, which can prematurely activate motility.

Quality Analysis: Evaluating sperm quality is crucial for selecting samples suitable for freezing. Parameters such as motility, viability, pH, and osmolality are assessed, often using advanced computerized systems to ensure accuracy.

Extender Formulation: Extenders are buffered solutions designed to prevent premature activation of motility and to provide an optimal environment for freezing. Common components include glucose, egg yolk, antioxidants, and CPAs like DMSO or glycerol. The choice of extender varies by species and specific requirements.

Freezing and Thawing: Sperm is typically loaded into French straws or cryovials and frozen at controlled rates before being stored in liquid nitrogen (-196°C). Thawing must be conducted rapidly in a water bath to ensure maximum viability. Figure 2.6 illustrates the sperm freezing procedures



Figure 2.6. Sperm freezing process: (A) trout sperm extraction by canulation, (B) dilution in a cryoprotectant extender, (C) loading in 0.5 cc French straws (insert with different straws, cryovials, and PVA powder for straw sealing), (D) freezing over a floating device in a styrofoam box containing N2l, (E) storage in a N2l tank, (F) female stripping, (G) sperm thawing in a water bath, and (H–J) fertilization (Fletcher & Rise, 2012)

4.3.2. Oocyte Cryopreservation

Unlike sperm, oocytes present significant challenges for cryopreservation. Their large size, complex structure, and limited permeability to CPAs make them highly susceptible to cryodamage. Issues such as chilling sensitivity, intracellular ice formation, and CPA toxicity are particularly pronounced. Moreover, the presence of multiple membrane layers and high lipid content further complicates the preservation process.

Recent research has focused on preserving oocytes at early developmental stages, where their structural simplicity may reduce susceptibility to cryodamage. Strategies include stepwise CPA removal to minimize toxicity, studies on chilling resistance, and the application of vitrification techniques. Vitrification, which involves ultrafast freezing with high CPA concentrations, offers a promising alternative by eliminating ice crystal formation. However, challenges remain in achieving uniform CPA distribution and minimizing toxicity.

4.4. Embryo cryopreservation

The cryopreservation of fish embryos, which aims to preserve both maternal and paternal genetic material, has significant potential for enhancing aquaculture reproductive management. Despite its promise, successful cryopreservation of fish embryos remains a challenge due to the biological complexities of embryos, such as their large size, multicompartmental structure, and limited permeability to cryoprotective agents (CPAs). These factors, combined with the presence of barriers like the yolk syncytial layer (YSL), hinder the effective distribution of CPAs and water throughout the embryo (Fig. 2.7, Hagedorn et al., 1997).



Figure 2.7. Turbot embryo at the tail bud stage showing the different envelopes and compartments: chorion (arrow), yolk syncytial layer (arrowhead), yolk sac (ys), perivitelline space (pvs), and embryo compartment (ec) (Hagedorn et al., 1997)

One major obstacle is the high water content in embryos, which can lead to ice formation and cellular damage during freezing and thawing. Early-stage embryos, which theoretically offer simpler structural properties for preservation, are highly sensitive to chilling and CPA toxicity, further complicating cryopreservation efforts.

Studies on chilling sensitivity in fish embryos have shown that early developmental stages are more vulnerable to low temperatures than later stages. Strategies to mitigate chilling injuries include modifying embryo structure and using protective substances such as antifreeze proteins (AFPs). These approaches have shown potential to enhance resistance to low temperatures but have not yet achieved consistent success.

The use of vitrification, a technique that eliminates ice formation through ultrafast freezing, has been proposed as a way to overcome these challenges. However, vitrification requires high concentrations of CPAs, which can be toxic and difficult to distribute evenly within the embryo due to its limited permeability. Various experimental techniques, such as increasing embryo permeability and improving CPA delivery systems, are being explored to address these limitations. Recent advancements include methods to bypass barriers like the YSL and improve CPA penetration. Techniques such as microinjection of CPAs or genetic engineering to enhance embryo permeability have shown promise. Additionally, the use of natural antifreeze proteins has demonstrated potential in reducing ice crystal formation and mitigating freezing-induced damage. Although these methods are still in experimental stages, they offer valuable insights into the future of embryo cryopreservation.

Overcoming the challenges of fish embryo cryopreservation will require interdisciplinary collaboration and technological innovation. Efforts are focused on enhancing cryoprotection at the cellular level and improving techniques for CPA delivery. Promising directions include the use of advanced laser technologies for creating temporary pores in embryos and developing genetically modified strains with enhanced resistance to freezing damage.

With continued research, fish embryo cryopreservation could become a reliable tool for aquaculture, supporting the preservation of genetic resources and promoting sustainable practices in fish farming.



4.5. Embryo cryopreservation

Cryopreservation technology has been developed for many fish species (Betsy et al., 2022):

- This technology can be used to preserve milt of the best age group brooder which can be used at any point of time in future.
- It can also eliminate inbreeding problem since cryopreserved spermatozoa can be easily exchanged between hatcheries.
- Using this technology, spermatozoa can be made available at any season of the year.
- It makes breeding possible during off-season.
- It synchronizes the gamete availability of both sexes leading to sperm economy.
- It simplifies broodstock management in farms.
- It helps in the production of viable and strong offspring by intra-species hybridization.
- It overcomes the difficulties arising due to the short time viability of gametes.
- It enables the genetic preservation of desired lines.
- It allows cross breeding at different times of the year.
- It helps in germplasm storage for genetic selection programs or conservation of species.
- Cryopreserved spermatozoa can help in the hybridization programs and genetic engineering research in fishes.
- It leads to many other avenues such as cryobanking of viable gametes as in the case of animal production and development of gene bank and genetic manipulation in fishes.

Cryopreservation represents a transformative tool in aquaculture biotechnology, offering significant benefits for genetic preservation, breeding programs, and biodiversity conservation. While challenges remain, particularly in embryo and oocyte preservation, ongoing advancements in cryoprotective methods, genetic tools, and interdisciplinary research hold promise for overcoming these barriers. Future developments will likely expand the scope and efficiency of cryopreservation, ensuring its broader application in aquaculture and beyond. Through continued innovation, cryopreservation is poised to play an integral role in supporting the sustainable growth of aquaculture and the preservation of aquatic biodiversity (Fletcher & Rise, 2012).

5. Ethical, Environmental, and Regulatory Considerations in Aquaculture Biotechnology

5.1. Ethical Concerns in Aquaculture Biotechnology

5.1.1. Animal Welfare in Genetic Modification

The ethical implications of genetic modification in aquaculture are profound, particularly regarding animal welfare. Genetic interventions, such as transgenesis and gene editing, often aim to enhance production traits like growth rates, disease resistance, or environmental tolerance. However, these modifications can inadvertently cause physiological stress or health complications. For instance, accelerated growth in transgenic fish may lead to skeletal deformities, reduced immune function, or altered metabolic rates. Critics argue that prioritizing productivity over welfare may compromise the ethical treatment of these organisms, raising questions about the balance between innovation and humane practices.

The confined nature of aquaculture systems further amplifies these concerns. Fish reared in such environments are often subjected to high stocking densities, leading to stress, susceptibility to



disease, and behavioral changes. Ethical considerations extend to whether genetically modified fish are more or less suited to thrive in such conditions compared to their wild counterparts. Developing welfare metrics specifically tailored for genetically altered aquatic species is essential to ensure their quality of life is not unduly compromised.

5.1.2. Ecological Integrity and Biodiversity

Beyond individual welfare, ethical concerns encompass the broader ecological impacts of biotechnological interventions. Introducing genetically modified or selectively bred species into aquaculture systems or natural habitats poses risks to ecological integrity. For example, transgenic fish with enhanced growth rates may outcompete native species for resources, disrupting local ecosystems and potentially leading to the decline or extinction of wild populations. These concerns underscore the moral responsibility of ensuring that biotechnology applications do not undermine the biodiversity and resilience of aquatic ecosystems.

The ethical debate also touches on human stewardship of biodiversity. While biotechnology can aid in conservation efforts, such as through cryopreservation of endangered species’ genetic material, it also raises questions about humanity’s right to alter genetic codes for economic or ecological purposes. Striking a balance between leveraging biotechnology for positive outcomes and preserving the natural evolutionary processes of aquatic species remains a key ethical challenge.

5.2. Regulatory Frameworks

5.2.1. Global Standards and Guidelines

The governance of biotechnological applications in aquaculture is a complex and evolving field. International organizations such as the Food and Agriculture Organization (FAO) and the Convention on Biological Diversity (CBD) have established frameworks to guide the safe and ethical use of biotechnology. These guidelines emphasize the precautionary principle, advocating for thorough risk assessments and monitoring before the approval and release of genetically modified organisms (GMOs) into aquaculture systems.

One key aspect of global standards is harmonizing regulations across countries to ensure consistency in safety measures and environmental protections. This is particularly important given the transboundary nature of aquatic ecosystems and the potential for escapees to impact neighboring nations’ waters. Collaboration among countries through treaties and agreements plays a crucial role in establishing uniform practices and mitigating risks.

5.2.2. National Regulatory Approaches

At the national level, regulatory frameworks vary widely, reflecting differing priorities, technological capacities, and societal attitudes toward biotechnology. Some countries, such as the United States and Canada, have robust systems for evaluating the safety and efficacy of genetically modified organisms, including extensive review processes involving scientific, environmental, and public health assessments. In contrast, other regions may lack comprehensive regulatory structures, leading to gaps in oversight and potential risks.

Regulatory approaches often include multi-step processes, beginning with laboratory testing and moving through controlled field trials before full-scale implementation. These processes aim to evaluate the environmental, economic, and social implications of new biotechnologies. Public consultation and transparency are increasingly recognized as critical components of regulatory frameworks, fostering trust and ensuring that decisions reflect societal values.



5.2.3. Safety Assessments and Approval Processes

Safety assessments are central to regulatory frameworks, providing a scientific basis for evaluating the potential risks of biotechnology applications. These assessments typically address several key areas:

Environmental Risks: Evaluating the likelihood of escape and the potential ecological impacts of GMOs, including competition with native species, hybridization, and habitat modification.

Human Health Risks: Ensuring that genetically modified fish intended for consumption are free from allergens, toxins, or unintended genetic effects that could harm consumers.

Ecosystem Monitoring: Implementing post-approval monitoring programs to detect and mitigate unforeseen impacts, ensuring long-term sustainability.

Approval processes often involve coordination among multiple agencies, including environmental, agricultural, and public health authorities. Rigorous scientific evaluations, combined with public input, aim to balance innovation with safety and ethical considerations.

5.3. Environmental Impacts of Aquaculture Biotechnology

5.3.1. Ecosystem Dynamics and Genetic Pollution

One of the most significant environmental risks of aquaculture biotechnology is genetic pollution, where genes from genetically modified or selectively bred species are transferred to wild populations. This can occur through interbreeding, leading to genetic homogenization and the loss of locally adapted traits in wild species. The long-term consequences of such genetic introgression include reduced resilience to environmental changes and diminished biodiversity.

The effects of domestication selection on the genetic and phenotypic characteristics of aquaculture animals lead to various potential environmental impacts upon release into nature. Fig. 2.8 summarizes the mechanisms responsible for such impacts within four categories: direct ecological effects, indirect ecological effects, direct genetic effects, and indirect genetic effects.

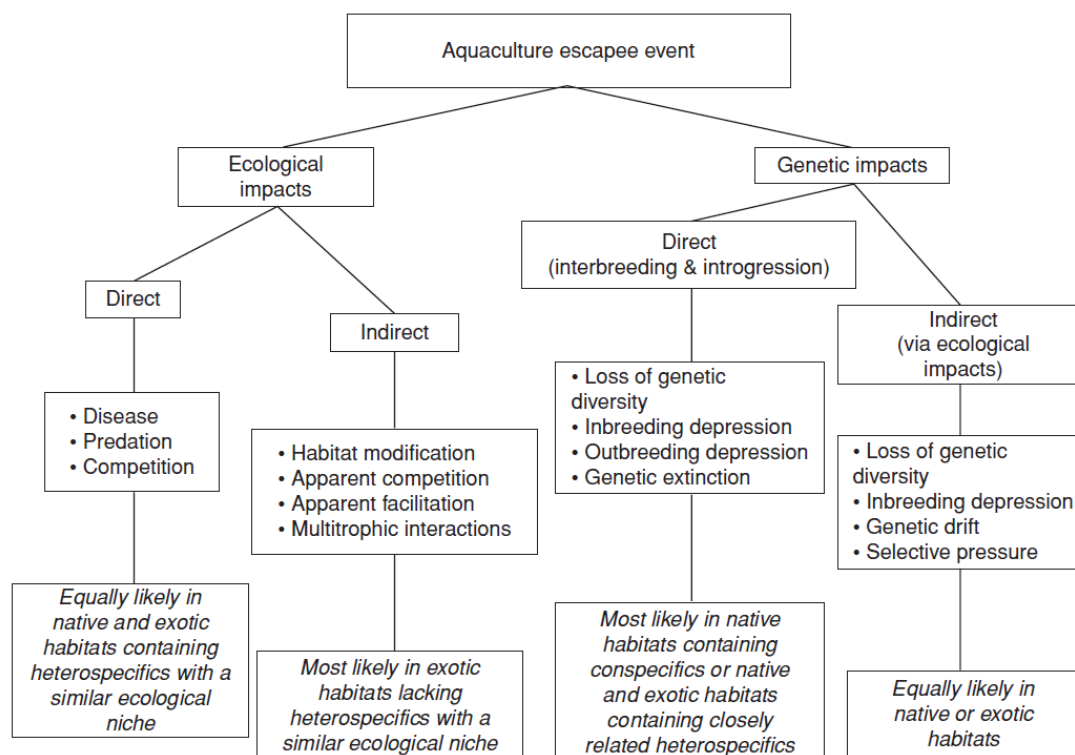


Figure 2.8. Possible environmental impact of aquaculture escapees



Aquaculture systems are particularly vulnerable to escape events, where farmed fish enter natural ecosystems. These escapees can outcompete wild populations for resources, introduce diseases, and disrupt food web dynamics. Mitigating these risks requires robust containment strategies, such as physical barriers, and developing sterile genetically modified fish to prevent reproduction in the wild.

5.3.2. Interactions with Wild Populations

Interactions between farmed and wild populations extend beyond genetic impacts. Transgenic fish with enhanced traits, such as faster growth or greater resistance to disease, may have ecological advantages over their wild counterparts. These advantages can lead to shifts in predator-prey relationships, altered competition dynamics, and changes in habitat use.

Research into the behavior and ecological roles of genetically modified fish is essential to anticipate and manage these interactions. Long-term ecological studies, combined with predictive modeling, can help identify potential risks and guide management practices.

5.3.3. Long-Term Sustainability

Ensuring the long-term sustainability of aquaculture biotechnology requires a holistic approach that considers ecological, economic, and social dimensions. This includes minimizing habitat destruction, optimizing resource use, and protecting wild populations. Advances in biotechnology, such as developing environmentally friendly feeds and improving waste management systems, can contribute to more sustainable aquaculture practices.

Monitoring and adaptive management are critical components of sustainable aquaculture. By continuously assessing the environmental impacts of biotechnological interventions and adjusting practices accordingly, stakeholders can balance productivity with ecological responsibility.

5.4. Balancing Progress and Responsibility

The integration of biotechnology into aquaculture offers immense opportunities for addressing global challenges such as food security and biodiversity conservation. However, this progress must be accompanied by a strong commitment to ethical principles, rigorous regulatory oversight, and proactive environmental stewardship. By fostering collaboration among scientists, policymakers, industry stakeholders, and the public, aquaculture can evolve in a manner that is both innovative and sustainable.

Ethical, environmental, and regulatory considerations are not merely obstacles to overcome but are integral to the responsible advancement of aquaculture biotechnology. Through careful planning, transparent decision-making, and ongoing research, the sector can realize its potential while safeguarding the well-being of aquatic ecosystems and the communities that depend on them (Fletcher & Rise, 2012).

Summary

Global warming has significantly disrupted the breeding cycles, growth rates, and survival of aquatic species. Rising water temperatures alter spawning times and metabolic rates, leading to mismatches with food availability and suboptimal conditions for larval development. Species like Atlantic cod and European perch are breeding earlier, resulting in reduced survival rates for their offspring. Additionally, elevated temperatures can decrease oxygen levels in water, stressing larvae and affecting juvenile development. While some species demonstrate genetic adaptations to cope



The Digital Blue Carrier for a Post-Carbon Future - Curriculum Innovations in Aquaculture [DiBluCa]”

2023-1-LT01-KA220-HED-000154247

with these changes, rapid environmental shifts often outpace the ability of populations to adapt, leading to long-term declines.

Aquaculture has embraced biotechnology to mitigate these challenges and enhance resilience in farmed species. Selective breeding programs focus on traits like heat tolerance, disease resistance, and growth efficiency. Genomic selection accelerates this process by using genetic markers to propagate desirable traits. For instance, Atlantic salmon have been bred to tolerate higher temperatures and hypoxic conditions, while genomic tools have been employed to develop disease-resistant strains of rainbow trout and other species.

CRISPR/Cas9 technology has emerged as a revolutionary tool in aquaculture, enabling precise and targeted modifications to fish genomes. This method allows for the enhancement of key traits such as growth, muscle quality, disease resistance, and environmental adaptation. For instance, genetic modifications in species like Nile tilapia and channel catfish have resulted in faster growth rates and improved muscle development by targeting the myostatin (*mstn*) gene. Similarly, CRISPR/Cas9 has been employed to enhance disease resistance in Atlantic salmon and grass carp by editing immune-related genes and pathogen recognition pathways.

In addition to improving individual traits, CRISPR has applications in sex determination and population management. Techniques such as creating sterile populations reduce ecological risks associated with escaped farmed fish. Despite these advancements, the technology is not without challenges. Off-target effects and ethical concerns surrounding genome editing, particularly in terms of animal welfare and ecological risks, necessitate robust regulatory oversight and further research.

Cryopreservation is another pivotal technology, offering solutions for genetic resource conservation and breeding efficiency. By preserving gametes and embryos at ultra-low temperatures, this technique supports biodiversity conservation and breeding programs across seasons and geographies. However, challenges such as chilling sensitivity and cryoprotectant toxicity, especially in oocytes and embryos, highlight the need for ongoing research to optimize protocols and improve success rates.

The integration of biotechnology into aquaculture raises profound ethical and environmental questions. The potential for genetically modified organisms (GMOs) to escape into natural ecosystems and interbreed with wild populations poses risks to genetic integrity and biodiversity. Regulatory frameworks at both national and international levels play a crucial role in addressing these concerns, emphasizing risk assessments, monitoring, and public engagement. Ethical considerations extend to animal welfare, particularly in ensuring that biotechnological interventions do not compromise the health and well-being of farmed species.

The future of aquaculture depends on balancing technological progress with sustainability. Innovations like CRISPR/Cas9 and genomic selection hold immense potential to enhance resilience and productivity. However, interdisciplinary collaboration, robust governance, and environmental stewardship are essential to minimize ecological impacts and ensure long-term viability. By prioritizing ethical practices and sustainability, aquaculture can play a pivotal role in addressing global food security challenges and conserving aquatic biodiversity.

This chapter underscores the urgency of addressing the interconnected challenges of climate change and aquaculture sustainability through innovative and responsible biotechnological solutions. By harnessing the potential of these advancements, the aquaculture industry can contribute to global efforts in biodiversity conservation, climate resilience, and food security.



References

- Angilletta, M. J., et al. (2004). "Thermal Adaptation of Ectotherms." *Nature*.
- Chevin, L. M., et al. (2010). "Adaptation to Climate Change." *Ecology Letters*.
- Durant, J. M., et al. (2007). "Trophic Match-Mismatch and Climate Change." *Ecology*.
- Fischer, J. R., et al. (2014). "Evolutionary Responses of Aquatic Species to Climate Change." *Nature Climate Change*.
- Heath, M. R., et al. (2014). "Climate Change and Fish Growth." *Fish and Fisheries*.
- Jorgensen, C., et al. (2017). "Local Adaptation of Atlantic Cod to Thermal Variation." *Journal of Evolutionary Biology*.
- Lynch, M., et al. (2014). "Evolution in Changing Environments." *Trends in Ecology & Evolution*.
- O'Reilly, C. M., et al. (2008). "Impacts of Climate Change on Fish Populations." *Science*.
- Parnesan, C. (2006). "Ecological and Evolutionary Responses to Recent Climate Change." *Annual Review of Ecology, Evolution, and Systematics*.
- Pörtner, H. O., et al. (2014). "Oxygen Supply and Temperature in Aquatic Ecosystems." *Nature*.
- Tompkins, E. M., et al. (2017). "Effects of Warming on Fish Breeding Patterns." *Global Change Biology*.
- Walther, G. R., et al. (2002). "Ecological Responses to Recent Climate Change." *Nature*.
- Gjøen, H. M., et al. (2018). "Aquaculture breeding programs for climate resilience." *Aquaculture Reports*.
- Houston, R. D., et al. (2018). "Selective breeding for disease resistance in aquaculture species: challenges and progress." *Fisheries Research*.
- Huntingford, F. A., et al. (2020). "The potential of selective breeding for climate resilience in aquaculture species." *Aquaculture*.
- Liu, Y., et al. (2020). "Genomic selection for heat tolerance in rainbow trout: A practical approach." *Journal of Fish Biology*.
- Vázquez, R., et al. (2018). "Genomic selection in aquaculture for disease resistance." *Aquaculture International*.
- Yang, Z., Yu, Y., Tay, Y. X., & Yue, G. H. (2021). Genome editing and its applications in genetic improvement in aquaculture. *Reviews in Aquaculture*, 00(1), 1–14. <https://doi.org/10.1111/raq.12591>
- Sankaran, G. B., & Mandal, A. (2024). Genetic improvements in aquaculture. *The Trout Journal of Atatürk University*, 2(1–2), 16–25. <https://doi.org/10.62425/tjau.1570599>
- Zhu, M., Sumana, S. L., Abdullateef, M. M., Falayi, O. C., Shui, Y., Zhang, C., Zhu, J., & Su, S. (2024). CRISPR/Cas9 technology for enhancing desirable traits of fish species in aquaculture. *International Journal of Molecular Sciences*, 25(17), 9299. <https://doi.org/10.3390/ijms25179299>
- Blix, T. B., Dalmo, R. A., Wargelius, A., & Myhr, A. I. (2021). Genome editing on finfish: Current status and implications for sustainability. *Reviews in Aquaculture*, 13(4), 2344–2363. [https://doi.org/\[CrossRef\]](https://doi.org/[CrossRef])
- Li, M., Dai, S., Liu, X., Xiao, H., & Wang, D. (2021). A detailed procedure for CRISPR/Cas9-mediated gene editing in tilapia. *Journal of Hydrobiology*, 848, 3865–3881. [https://doi.org/\[CrossRef\]](https://doi.org/[CrossRef])



The Digital Blue Carrier for a Post-Carbon Future - Curriculum Innovations in Aquaculture [DiBluCa]”

2023-1-LT01-KA220-HED-000154247

Wargelius, A., Leininger, S., Skaftnesmo, K. O., Kleppe, L., Andersson, E., Taranger, G. L., Schulz, R. W., & Edvardsen, R. B. (2016). Dnd knockout ablates germ cells and demonstrates germ cell independent sex differentiation in Atlantic salmon. *Scientific Reports*, 6, 21284. [https://doi.org/\[CrossRef\]](https://doi.org/[CrossRef]) [PubMed]

Betsy, C. J., C, S., & Sampath Kumar, J. S. (2022). Cryopreservation and its application in aquaculture. In *Cryopreservation and Its Application in Aquaculture*. IntechOpen. <https://doi.org/10.5772/intechopen.99629>

Fletcher, G. L., & Rise, M. L. (Eds.). (2012). *Aquaculture biotechnology*. Chichester: Wiley-Blackwell.

Hagedorn, M., Hsu, E., Kleinhans, F. W., & Wildt, D. E. (1997). New approaches for studying the permeability of fish embryos: Toward successful cryopreservation. *Cryobiology*, 34(4), 335–347.